

GENERAL LAB RULES  
PALEOETHNOBOTANY LABORATORY  
UNIVERSITY OF MINNESOTA  
1989

Please be neat and keep areas around microscopes clean, as we have to share equipment and space. Wash counter space from time to time to keep work areas and data sheets clean.

Keep the sample you're working on, your sorting box, your data sheets, brushes, tweezers, etc. in your own personal drawer.

When storing unfinished samples, make sure they are well labelled, and covered.

It is best not to turn illuminators off except when you are done for the day, this saves on bulbs and mechanical failures.

Feel free to bring in tapes, or play the ones in the lab. Please don't ever play them loudly (even on week-ends). Do not take any of the resident tapes out of the lab.

Please keep conversations quiet, work may seem mindless, but it is important to pay attention to what you're doing, and try not to disturb or distract others. Be especially mindful on days when there are many workers in the lab.

Please hang up coats, and put book bags under desks, or out of the way. Keep other personal belongings in your drawer. Throw newspapers in the recycling box. Bottles and cans for recycling go under the sink.

At the beginning of each quarter fill out a schedule of the hours you wish to work. It is best to work for more than one hour, but less than 3 at a microscope at one time. A total of four hours a day at the microscope is the allowed maximum. If you need to work for more than three hours at the microscope in one day, sign out and take a break. This is very important, and these rules cannot be broken. Remember to stick to your schedule, and if you cannot come in, please call and let us know.

Everyone working on sorting and ID's must sign in on the sign-in sheet.

Keep time cards up to date, and remember to get Dr. Hastorf's signature and turn them in to Terri V. in Anthropology on time.

Time cards should include time spent sorting, working on identifications, stamping boxes, weighing samples, and mandatory lab meetings; they should not include time spent eating lunch, talking on the phone, or optional lab meetings or presentations.

Please fill in only the actual hours you work, remember it takes time to get to and from classes.

There is a hot plate and refridgerator available for everyone to use. Please clean up after yourself, and keep clutter away from microscope stations near the sink.

Please use the telephone sparingly, no long distance calls.

Remember to cover your microscope when you are finished, and to turn out the lights, turn off the humidifier (if it's on), and lock the door if you are the last to leave.

If there are any building problems tell them downstairs in the main office.

Shepherd labs is locked after about 4:30 weekdays and on week-ends.

There is no smoking in the lab.

## BOLIVIAN MATERIAL FROM DR. ALAN KOLATA'S WILA JAWIRA PROJECT

This includes material from excavations at Lukurmata, and all parts of Tiwanaku. Flotation samples from these sites were taken starting in 1986 (at least), and continue through the present. His excavation and recording strategy centres around an infinitely expandable grid, noted as UNIDADES North and East of a single datum. Each samples can have the following information:

F# = Flot number, unique w/in our lab. Starts at 4000  
SITE = site, LKM= Lukurmata, TIW= Tiwanaku,  
CUADRA = Can be numbers or letters, is sub-section of site  
NIVEL = Level  
SPECIMEN # = Bag number which is unique to each site  
FECHA = Date excavated, watch out for order of month and date  
UNIDAD = Unit (nearly always given as Nxxxx Exxxx)  
RASGO = Feature number, sometimes w/description  
EXCAVATOR = Initials of head of dig team

Other information to be filled in on our laboratory analysis forms:

FLOTVOL = Volume of sample in litres, found in flot log  
COLLTYPE = whether sample is BULK (101) or PINCH (102). Check  
flot log for details. Screen material (1/4") is 201  
LFWT = Combined total of Light Fraction, both carbon and remains  
PICKWT = Weight of carbon sorted out of the sample  
CC = Cultural Context of sample. This is found in the field notes  
SCREEN Y/N = Do we have some material from the screen for this  
location?

#### Filling in forms:

Select a provenience, and check for all samples that were taken. For each sample there may be a heavy fraction, if so, it should be attached to the light fraction bag. Screen material have the same provenience number, and take care as there can be more than one for each provenience, or none at all. A data sheet should be started for the flot and the associated screen. Remember to use a (sharp) pencil. Check also for a possible "FLOT PESADO" which is unsorted, <2mm heavy fraction material.

On the flot sample form the following should be filled out by the sorter:

SITE, CUARDA, NIVEL, SPECIMEN#, FLOT#, UNIDAD, RASGO, FLOTVOL, COLLYPE, CC (if you can figure this out), SCREEN Y/N, Sorter's initials and date sample was started in the correct spot, IE: if you are doing the LIGHT fraction and /or the HEAVY fraction. On the back fill in the provenience and flot number again, as well as the initials of the excavator and the date excavated. Fill in what percent of the sample was sorted, and if you split any portion of the sample write in the amount it weighed. EG: LF>2mm =100%, LF<2mm and >1.18 =50% (1.2g), LF<1.18 and >0.5 =12.5% (2.1g), and LF<0.5 =1.25% (1.5g). Check off and write in qualitative estimate of amounts of fish scales in the fractions you have sorted (if we are not currently picking them out). Also draw rough sketch of any land snails, with an estimate such as "lots" or "few". Don't bother to count them. Under "comments" add any other information on the tags.

#### Processing flotation samples:

Empty the light fraction into the geologic sieves, replace lid, and shake carefully. If necessary, carefully loosen roots with probes. Save plastic bag, or if cut open, the portion with labels.

Gently shake material from the 2mm screen into a glass dish. It is best to sort all >2 material in glass in case it is needed for radiocarbon dating. For this same reason, it should not be touched with your hands. Material from the bottom sieve can be sorted in a cardboard box. Make sure to keep a label in each of the subsamples

If there is a lot of small material and dust in a sample you may also want to use the 1.18mm, 0.5mm and 3.0mm sieves to make it easier to sort.

NB: Check the section of this document for sorting strategies (p9-11), especially if you are going to do less than a "complete" sort.

Sort each fraction twice, systematically.

Remove all carbon from each fraction, except wood that is <2mm.

Remove all bone and fish scales, whether charred or not (if you are doing a complete sort).

Put all >2 items in a used vial, and all remains in a used box. If you are not working on identifications put all <2mm material into another used vial. If you are working on identifications put different seeds into separate gelitan capsules as you sort. If possible separate unidentifiable seeds from lumps. Keep the capsules in the holder, and avoid contact with your hands as much as possible. When you have finished a sample take all capsules and place them in a clean vial or box, be sure to include a label.

Turn in remains, carbon, bones, empty plastic bag or labels, data sheet(s), and the unsorted heavy fraction and screen bots.

#### Weighing samples:

After samples are checked and are ready to be put away they must be weighed, and the boxes stamped.

Weight the entire light fraction, including identified material and remains. Subtract the weight of the gelitan capsules, plastic bags, and vials.

Plastic bags = 2.5g

Gelitan capsules = 0.1g

1 Vial = 6.4g

2 Vials = 12.7g

3 Vials = 19.1g

4 Vials = 25.5g

5 Vials = 31.9g

6 Vials = 38.3g

This value is filled in as LF WEIGHT.

Then weigh all the picked material (carbon and bones), fill this in as PICKED WEIGHT.

After weighing remove all capsules with coloured labels from the light fraction, heavy fraction, and the screen bots. In the upper right hand section of the form, use the appropriate stamp to indicate that these materials have been removed, and put them in the boxes with others that have been removed from other samples. Yellow tags are for bones, pink for fish scales, blue for unknown seeds, and orange for artefacts.

Return the material to the box, being careful that the lid fits securely, and that none of the capsules are squished. On the upper right hand corner and one end stamp the provenience, year of excavation, and the flot number.

For Peruvian UMARP samples:

EG: 1-477/2	2=1-1-3-2-5/7
1986	1983

# 2553

# 1279

For Bolivian Wila Jawira samples:

EG: SITE TIW                    UNIDAD N788 E5125  
CUADRA AK-E                    RASGO 1  
NIVEL 3                         FLOT # 4023  
SPECIMEN 15223                 YEAR 1989

See the examples on the shelves. Then put each sample in order by provenience (or by SITE and SPECIMEN # for Bolivian material), within each of the three box sizes.

--PLEASE WRITE NEATLY ON DATA SHEETS, REMEMBER  
SOMEONE ELSE WILL HAVE TO READ THEM--

## SORTING STRATEGIES FOR ARCHAEBOTANICAL MATERIAL IN THE LAB

Because time and money are always in high demand in the lab there are several different strategies that can be used when sorting and identifying archaeobotanical material in the lab. Other considerations are the goals of the study at hand, the quality of the collection and recovery techniques used to retrieve botanical material, and the overall quality of archaeological information available for the interpretation of the materials.

Below are schemes devised especially for flotation samples, where the study of domesticates is the main focus.

### Strategy 1: Complete sort

In the best of all possible worlds it is nice to be able to sort out and identify all useful material from a sample. It is especially desirable because a single float sample is already only a small sample of any given archaeological context, and one wants as complete a picture as possible. In our case, one would sort out, and identify all charred material, except <2mm wood, which is usually unidentifiable. All bones and other animal and artifactual materials are pulled out and given to appropriate specialists.

This type of strategy gives RATIO level data, with exact counts (and/or weights) entered onto the computer. Descriptive statistics such as RELATIVE PERCENTAGES, DENSITIES, UBIQUITIES, and DIVERSITIES can be generated from this type of data.

This strategy is the most labor intensive, and can be a waste of time when you sort past the point of diminishing returns, ie, you get the exact same values by sorting entire sample that you would by making estimates based on some fraction of the whole (50%, 25%, etc).

### Strategy 2: Sample splitting

In this strategy time is saved by splitting (by weight) some or all of the sample. It is usually done at one of the smaller fractions separated by the geologic sieves, eg, 100% of the material that is >2mm is sorted, while 50% of all material <2mm is sorted and all counts of the identified specimens are doubled. The decision to split a sample should be based on the following guidelines. The average amount of time spent on a sample is about 2 1/2 hours, including sorting and identifying light and heavy fractions, as well as material recovered from the sieves in the field. The two main factors that should be considered are both the volume of the sample, and the density of the seeds. The desired amount of material to sort from each size fraction of the sample is enough to fill one of the sorting trays (in a thin layer, as when ready for sorting). If a brief scan of even this amount appears to contain hundreds of seeds, it should be split again. A rule of thumb that has proven effective for the 1986 Pancán (Peru) material was never to let the sorted portion fall below 1.0g or 12.5% (3 times though the sample splitter). In

these samples it was found that this was approximately the point of diminishing returns for very dense samples such as those from burnt stores of crops, where seeds and tuber densities per 6-liter of soil averaged in the thousands. That is, if at least these 12.5% or 1.0g of each size fraction was sorted the estimates of total densities and taxa diversity were found to be insignificantly different than if the whole sample had been sorted. It should be noted on the form which fractions were split, what percentage was sorted, and the weight of the material prior to sorting. Of course, special circumstances may occur, and less may be sorted without losing accuracy.

Trials with a 0.3mm geologic sieve show that very, very few seeds will pass through this mesh size. Another time saving measure in dusty samples is not to sort the material that is less than 0.3mm. If bones and fish scales are too numerous, they can be left in remains and notes of their occurrence and/or abundance can be put on the data sheet. If very small lumps are overabundant one can leave those <1.18mm (with no distinctive characteristics, such as a surface) in the remains.

As with the complete sort, one gets RATIO level data, and can generate RELATIVE PERCENTAGES, DENSITIES, UBIQUITIES, and DIVERSITIES. Because actual counts are estimated this type of data can be used in comparison with that of Strategy 1 with no conversion.

This method is a good time saver, especially for samples that are quite homogenous. Drawbacks are that diversity may be lost, and rare species are either missed or overrepresented.

### Strategy 3: Scanning with estimates and removal of special taxa

In this scheme only domesticates are removed from the remains. The samples are scanned and the weedy species are recorded as "common", "occasional", or "rare" (corresponding to values of 3, 2 and 1, respectively). Each of these categories is based on a visual estimate of the number of each different taxon. Counts corresponding to each category should be determined so that all workers are making the same estimates.

This will result in ORDINAL level data, and UBIQUITIES and DIVERSITIES are possible, as are some other nonparametric statistics such as Spearman's Rho.

Remember the size of the pre-flot soil sample must be taken into consideration, and that this should only be done when samples were originally similar in size. If any sorter is unfamiliar with any taxon it too must be removed for identification. For this reason this strategy is less time-efficient with beginning sorters. Simple routines on the computer can be used to assign data form strategies 1 and 2 to the same format as these, eg "if Verbena=1 then Verbena=1; if Verbena >1 and <5 then Verbena=2; if Verbena >5 then Verbena=3.

### Strategy 4: Scanning

In this scheme no specimens are removed from the sample. Instead, a complete scan is made of the sample, and the presence of taxa is marked off the data sheet.



This results in NOMINAL level data, from which can be used to generate UBIQUITIES and DIVERSITIES, and any scheme that only requires presence/absence data.

The computer can again be used to assign data to presence/absence level very simply (usually as 1 and 0), eg, if Verbena >0 then Verbena =1, etc. This routine is the fastest, and can be useful when only a rough sketch is needed from the dataset, often as a preliminary method to further detailed work. The drawbacks can be the loss of quantitative data, and requires all workers to know all taxa.

CULTURAL CONTEXT CODES FOR USE WITH 1986-1989 WILA JAWIRA  
MATERIAL:

Surface and sub-surface modern disturbance:

- 000 General surface collection
- 010 Humus root zone, do NOT combine in analysis
- 020 Plowed surface collection
- 021 Plowed surface-shovel scraped
- 030 Fallow/harvested surface collection
- 031 Fallow/harvested surface-shovel scraped
- 040 Natural/wild surface collection
- 050 Plough zone
- 060 Excavated surface collection
- 070 Modern wall or rock pile
- 080 Humus root zone, okay to combine in analysis w/level below
- 090 Modern burned area
- 091 Modern animal burial

Wall:

- 100 Possible wall
- 110 Rock wall, unmortared
- 120 Pirka wall
- 121 Outside supportive lip
- 122 Inside supportive lip
- 125 Rock wall, single course wide
- 126 Fill/mortar from wall
- 130 Dressed stone wall
- 140 Rock wallfall
- 141 Adobe wallfall
- 150 Wallfall, do NOT combine in analysis
- 160 Wall trench fill
- 161 Wall trench
- 162 Wall plaster, slumped off
- 163 Wall plaster facing
- 170 Retaining wallfall
- 180 Wallfall, okay to combine in analysis w/level below
- 190 Adobe/ mudwall
- 191 Stone foundation of adobe wall
- 192 Adobe and rock wall

Midden; culturally deposited:

- 200 Low density midden
- 201 Low density midden-primary deposition
- 202 Low density midden-secondary deposition
- 210 Medium density midden
- 211 Medium density midden-primary
- 212 Medium density midden-secondary
- 220 High density midden
- 221 High density midden-primary
- 222 High density midden-secondary
- 230 Low density midden with ash
- 231 Low density midden with ash-primary
- 232 Low density midden with ash-secondary

240 Medium density midden with ash  
241 Medium density midden with ash-primary  
242 Medium density midden with ash-secondary  
250 High density midden with ash  
251 High density midden with ash-primary  
252 High density midden with ash-secondary  
260 Plough zone derived from midden  
270 Midden interspersed w/natural deposition  
280 Midden interspersed w/wall slump  
291 Cut below midden deposit  
298 Midden-details unspecified  
299 Midden-stratified

Cultural Surfaces; "use" surfaces and their deposits:

300 Surface, not clearly compact  
301 Surface inside structure  
302 Surface outside structure  
310 Occupation zone, matrix deposited during use  
311 Occupation zone, matrix deposited during use-inside  
312 Occupation zone, matrix deposited during use-outside  
313 Dense occupation zone  
314 Occupation zone w/disturbed, burnt "jacal"  
320 Activity area  
321 Metal processing area  
322 Food processing area  
323 Ceramic production area  
324 Storage area-burnt in situ  
330 Floor contact (material on floor surface)  
340 "Crusty", compact surface  
341 Cut associated w/compact surface  
342 Compact surface inside structure (true floor)  
343 Compact surface outside structure  
344 Clay floor inside structure  
350 Paved floor  
351 Paved floor inside structure  
352 Paved surface outside structure  
360 Rock subfloor/ cobble drain construction  
370 Occupation zone with roof or wallfall  
380 Plough zone derived from occupation zone  
390 Possible occupation zone

Features; culturally deposited:

400 General  
410 Pit fill  
412 Pit fill-midden  
413 Pit fill-gravel  
414 Pit fill-natural matrix w/artifacts  
415 Pit fill-ash  
416 Pit fill-clay  
417 Pit with camelid bones  
418 Pit with cuy bones  
420 Hearth (in situ burned area w/well defined limits  
421 Hearth cut

422 Ephemeral in situ burned area (not associated w/clear cut)  
423 Hearth-stone and adobe lined  
424 Burned area of floor-inside  
425 Oven  
430 Subfloor drainage canal  
440 Stairway  
450 Other firing feature  
451 Burned clay concentration-NOT in situ  
460 Ash deposit (not a clear lens or pit)  
470 Posthole fill  
471 Cut of posthole  
480 Stone fill (cultural) purpose unclear  
490 Possible feature  
498 Fill from inside of ceramic vessel  
499 Fill from bell-shaped pit

Burials:

500 Burial in subfloor-primary  
510 Burial in subfloor-secondary  
520 Burial in midden-primary  
530 Burial in midden-secondary  
540 Burial in patio-primary  
550 Burial in patio-secondary  
560 Burial in wallfall  
580 Animal Burial  
591 Cut below burial  
592 Burial in natural matrix w/artifacts  
593 Burial in capped, collared cist tomb  
594 Burial in belled-pit tomb  
598 Burial, details unspecified

Fill; purposefully deposited, but that contains artifacts  
unrelated to location:

600 Human dumped natural matrix w/artifacts  
601 Rapid-water deposited matrix w/artifacts  
602 Long-term erosion-deposited matrix w/artifacts  
603 Decomposing bedrock w/artifacts  
610 Midden used as fill  
620 Cultural fill  
621 Cut below fill  
622 House construction fill inside house  
623 House construction fill under house  
624 Rocky fill (purposeful)  
625 Gravel fill (purposeful)  
650 Naturally deposited soil, sterile  
670 Culturally deposited matrix w/few artifacts  
690 Possible fill  
699 Gravel fill as foundation of raised field

Lenses; thin deposits (cultural deposits, natural deposits or reworking of cultural deposits):

- 700 Ash lens, grey-white ash
- 710 Gravel lens
- 720 Charred lens-black
- 730 Natural matrix lens, water deposited
- 740 Organic stain

No good evidence for interpretation of depositional history:

- 900 Undifferentiated soil
- 910 Undifferentiated rock
- 920 Locus unexcavated
- 999 Mixed locus or information lost or incorrect-check notes before analysing